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HELICOBACTER PYLORI AS A RISK FACTOR FOR THE DEVELOPMENT OF METABOLIC SYNDROME AND GALLSTONE DISEASE

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HELICOBACTER PYLORI КАК ФАКТОР РИСКА РАЗВИТИЯ МЕТАБОЛИЧЕСКОГО СИНДРОМА И ЖЕЛЧНОКАМЕННОЙ БОЛЕЗНИ

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To study the *Helicobacter pylori* (*H. pylori*) effect on gallstone disease development and metabolic syndrome (MetS) and its components, the main factors of biliary stone formation. The study included 143 patients with cholelithiasis, of which 88 (61.5 %) were infected with *H. pylori* and 55 (38.5 %) were not. All subjects were assessed for MetS and its components' severity. A high prevalence of metabolic disorder and MetS was revealed in patients with *H. pylori*. The results obtained suggest that *H. pylori* promotes gallbladder calculi development by affecting patients' metabolic profile.

Keywords: gallstone disease, Helicobacter pylori, metabolic syndrome

Изучено влияние *Helicobacter pylori* (*H. pylori*) на развитие желчнокаменной болезни (ЖКБ), а также метаболического синдрома (МС) и его компонентов – основных факторов, влияющих на формирование желчных конкрементов. В исследование было включено 143 пациента с ЖКБ, из которых 88 (61,5 %) инфицированы *H. pylori* и 55 (38,5 %) не инфицированы. Среди всех испытуемых была проведена оценка выраженности МС и его компонентов. В результате была выявлена большая распространенность метаболических нарушений и МС у пациентов с *H. pylori*. Таким образом, полученные результаты дают основание полагать, что *H. pylori* способствует развитию конкрементов в желчном пузыре путем воздействия на метаболический профиль пациентов.

Ключевые слова: желчнокаменная болезнь, Helicobacter pylori, метаболический синдром

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AH	– arterial hypertension	MV	– mean value
BMI	– body mass index	NK-cells	– natural killer cells
DBP	– diastolic blood pressure	OR	– odds ratio
DM	– diabetes mellitus	s.d	– standard deviation
GSD	– Gallstone disease	SBP	– systolic blood pressure
<i>H. pylori</i>	– <i>Helicobacter pylori</i>	T4SS	– type IV secretion system
HDL-C	– high-density lipoprotein cholesterol	TC	– total cholesterol
HMG-CoA	– 3-hydroxy-3-methylglutaryl coenzyme A	TG	– triglycerides
IL-1, -6, -8, -10	– interleukin -1, -6, -8, -10	UDCA	– ursodeoxycholic acid
JIS	– Joint Interim Societies	WC	– waist circumference
LDL-C	– low-density lipoprotein cholesterol	95 % CI	– 95 % confidence interval
MetS	– metabolic syndrome		

Gallstone disease (GSD) is one of the most common gastrointestinal tract pathologies, often resulting in complications in cholecystitis, pancreatitis, and secondary infections. It is also associated with digestive system cancer. According to epidemiological data, gall bladder calculi are found in 10–20 % of developed countries' inhabitants [1]. At present, risk factors for cholelithiasis development are well studied. The main ones determining the disease formation and progression are metabolic syndrome (MetS) and its components, which include overweight, diabetes mellitus (DM), arterial hypertension (AH), and dyslipidemia. Recently, however, researchers have been interested in studying the *Helicobacter pylori* (*H. pylori*) role in developing GSD and metabolic disorders.

H. pylori is a gram-negative microaerophilic spiral bacterium with flagella. It colonizes the stomach mucous membrane in half of the world's population, exposing them to a high risk of gastritis, peptic ulcer, and gastric adenocarcinoma. Over the past few years, numerous studies have confirmed the linkage between cholelithiasis and *H. pylori* infection [2, 3]. This microorganism is also found in tissue samples of the bile, calculi, gallbladder, and liver. *H. pylori* virulence factors include *vacA*1, *iceA*1, *babA*2, *cagA*, and *cagE* genes, which play different roles in GSD pathogenesis [4]. Researchers have found that *vacA*1, *iceA*1, and *babA*2 are the most predominant genotypes in groups of patients with cholangiocarcinoma and GSD. Both *cagA* and *cagE* can be potential markers for predicting hepatobiliary disease severity [5]. A link between *H. pylori* infection and metabolic disorders has also been proven [6]. *H. pylori* eradication leads to a decrease in metabolic changes associated with a decrease in waist circumference (WC), glycemic and lipid profiles [2, 3]. It can be assumed that GSD high prevalence among the population can be linked with *H. pylori* infection rapid growth and its effect on the metabolic profile. In this regard, the study of this microorganism's role in MetS development in GSD patients is fundamental.

Material and Methods. A total of 143 patients with GSD aged 40 to 65 years were examined, with 110 (77 %) women and 33 (23 %) men among them. The participant's average age was 55.2±7.51 years. Among 143 patients with cholelithiasis, *H. pylori* infection was detected in 88 (61.5 %). All patients gave written informed consent to participate and were examined following the World Health Organization Helsinki Declaration principles and bioethics rules. The work took into account the following data: anamnestic (the presence of AH and DM), anthropometric (height, body mass, body mass index (BMI), WC) and biochemical (total cholesterol (TC), high-density lipoprotein cholesterol (HDL-C), low-density lipoprotein cholesterol (LDL-C), triglycerides (TG), glucose). In addition, MS verification was carried out based on the JIS criteria (Table 1).

Gallbladder stones were detected using ultrasound of the abdominal organs. The material for a cytological study to detect *H. pylori* infection was Romanovsky-Giemsa

stained smear-prints of biopsy samples obtained by esophagogastroduodenoscopy from mucous membrane sections of the stomach antrum.

Statistical analysis was carried out per the goal. The Kolmogorov-Smirnov test was used to assess the distribution normality of variables. $MV \pm s.d$ and 95 % CI were calculated for numerical variables. The U-Mann-Whitney test was applied, taking into account sample normal distribution. Comparison of the two groups' qualitative data was carried out using contingency tables of Fisher's exact test. The SPSS 22.0 program (SPSS Inc, USA) for Windows (Microsoft Corporation, USA) was used for statistical analysis, while $p < 0.05$ was considered statistically significant.

Table 1

MS JIS, 2009 Criteria

1	WC > 94 cm (men), > 80 cm (women)	Any 3 of the listed
2	Systolic blood pressure (SBP) > 130 mm Hg and diastolic blood pressure (DBP) > 85 mm Hg *	
3	TG > 1.7 mmol/L **	
4	HDL-C < 1.0 mmol/L (men), < 1.3 mmol/L (women) ***	
5	Fasting glucose > 5.6 mmol/L ***	

* Or for previously diagnosed AH treatment.
** Or for this disorder-specific treatment.
*** Or previously diagnosed type 2 DM.

Results and Discussion. A prevalence assessment of metabolic disorder components was made in conformity with the research goal. It was found that according to JIS-2009 criteria (Joint Interim Societies, 2009), GSD patients infected with *H. pylori* are more likely to have all MetS components compared with GSD patients without *H. pylori* infection (Fig.). The most pronounced differences were observed in WC criteria > 94 cm in men, and > 80 cm in women (76.1 % in *H. pylori*-positive cases versus 49 % in *H. pylori*-negative ones, $p < 0.05$), TG > 1.7 mmol/L (56.8 % in *H. pylori*-positive cases versus 38.1 % in *H. pylori*-negative ones, $p < 0.05$) and HDL-C < 1.0 mmol/L (men), < 1.3 mmol/L (women) (30.6 % in *H. pylori*-positive cases versus 10.9 % in

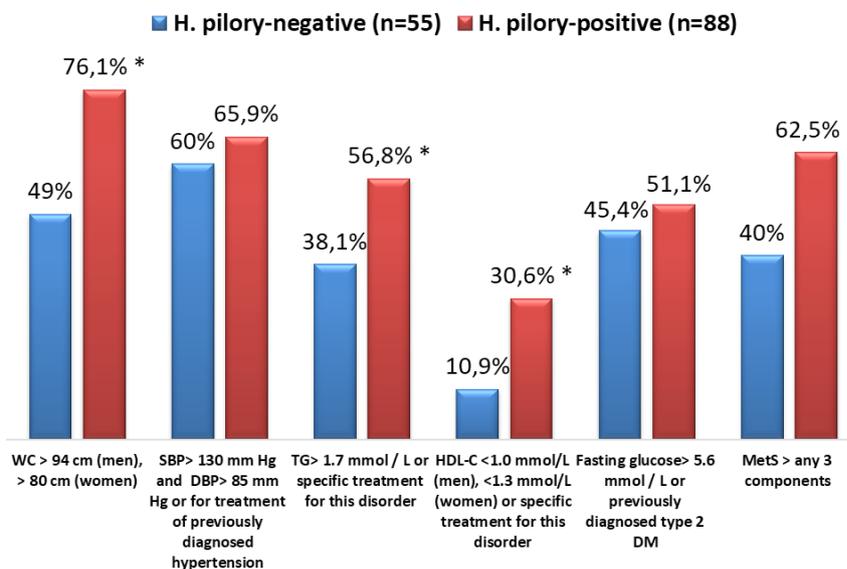


Fig. Metabolic disorder prevalence, based on the JIS-2009 criteria in individuals with GSD depending on *H. pylori* infection (* – Fisher's exact test $p < 0.05$)

H. pylori-negative ones, $p < 0.05$). A patient with three or more MetS components, according to JIS-2009 criteria, was recognized as having MetS. Thus, the study results demonstrate a higher MetS prevalence in GSD patients with H. pylori infection compared with GSD individuals without it (62.5 % in H. pylori-positive versus 40 % in H. pylori-negative, $p < 0.05$).

We assessed the average rate of anthropometric measurements and blood serum chemistry (Table 2). It was found that individuals with H. pylori infection are more prone to obesity compared to uninfected ones, as they have higher average weight, BMI, and WC rates. Also, these patients had more pronounced lipid metabolism disorders. They often had high TC, LDL-C, and TG levels and lower HDL-C levels compared with uninfected patients. Also, among GSD patients infected with H. pylori, more pronounced carbohydrate metabolism disorders compared with H. pylori-negative patients. With the help of U-Mann-Whitney test, it was found that in GSD patients H. pylori infection statistically significantly affects weight gain ($U=1334.5$; $Z=-4.506$; $p=0.001$), BMI ($U=1233.5$; $Z=-4.924$; $p=0.001$), WC ($U=1333.5$; $Z=-4.511$; $p=0.001$), an increase in plasma levels of TG ($U=1823$; $Z=-2.477$; $p=0.013$) and a decrease in the level of HDL-C ($U=1791$; $Z=-2.610$; $p=0.009$).

76.66 % of patients with cholelithiasis who were infected with H. pylori [7]. A meta-analysis by D. Zhou et al. found a trend towards higher H. pylori infection among GSD patients than in control groups [3]. Another 2018 meta-analysis, which included 18 studies, found that H. pylori infection was closely connected with an increased risk of developing cholelithiasis [2]. It is reported that H. pylori has an effect on the entire hepatobiliary system since several researchers have managed to obtain the H. pylori bacterium from bile, calculi, and tissue samples of the gallbladder and liver in patients with GSD.

Gallstones are known to be a reservoir for bacteria. At the same time, mucus production facilitates their adhesion, and β -glucuronidase and phospholipase form a surface for colonization. In turn, gallstones enriched with bacteria exacerbate disease severity, depending on the bacterial composition. At the same time, dense microorganism clusters in biofilm attached to the mucous membrane surface and calculi are more resistant to antibacterial drugs [8]. By modifying the lipopolysaccharide lipid component, H. pylori evade recognition by the host immune system. Proteomes can trigger another process of immune intervention on biofilm containing H. pylori.

Biofilm is a reservoir for a variety of microorganisms, in particular for H. pylori. It promotes genetic exchange between them. Amplification of antibiotic-resistant genes spreads in biofilms due to horizontal transfer of genes, which are integrative to conjugative elements [9]. In addition, stressful conditions can contribute to mutations and the emergence of antibiotic-resistant strains. In vitro studies have shown that the *cagA* and T4SS genes are involved in biofilm formation with H. pylori [10]. It is important to note that H. pylori CagA is an oncoprotein produced in the bacterial cytosol and then penetrates the host cell cytosol using a type IV cell secretion system (T4SS). CagA regulates autophagy, reduces epithelial cell apoptosis by interacting with tumor suppressors such as p53 [11], stimulates an inflammatory response via the c-Met-PI3K/Akt-mTOR signaling pathway, [12] and is a marker for predicting hepatobiliary disease severity [5]. An interesting fact is that H. pylori is a cholesterol auxotroph since it extracts lipids from host membranes to incorporate it into its outer shell using the cholesterol- α -glucosyltransferase enzyme. Cholesterol glucosylation and extraction from host cells lead to lipid raft destruction and/or a change in the membrane structure associated with immunity evasion and bacteria resistant to treatment [13]. Besides, there is a distinct shift in the distal intestinal tract microbiota resulting from H. pylori infection development. Undoubtedly, the imbalance between the gastrointestinal tract microbiota and the host contributes to immune, inflammatory, and metabolic disorder development [14].

We found a positive relationship between H. pylori and MetS in GSD patients for whom both MetS and its components were much more common (Fig., Table 2). Many researchers have confirmed the link between H. pylori carriage and MetS. So, a meta-analysis, which included 18 studies with 27,544 participants, demonstrated H. pylori and MetS correlation (odds ratio (OR)=1.34, 95 % CI=1.17–1.53, $p < 0.01$) and significant difference in glucose, HDL-C, BMI, TG and blood pressure between infected and uninfected groups of patients [6].

H. pylori colonising the crypts and gastric mucosa inevitably leads to chronic gastritis with mild chronic inflammation. Although H. pylori does not enter the bloodstream, the infection's long-term effects are induced by circulating cytokines, acute phase proteins, and other mediators. In addition, H. pylori virulence factors induce inflammatory proteins and cytokines such as C-reactive protein, tumor

Table 2

Results of anthropometric measurements and blood counts in GSD patients depending on H. pylori infection

Indicator		Mean value (MV) \pm standard deviation (s.d)	95 % CI (confidence interval)	Mann-Whitney U-test	Z	p
Age	1	54.6 \pm 7.2	52.6–56.6	2186.5	-0.971	0.332
	2	55.6 \pm 7.6	54–57.2			
Weight	1	69.1 \pm 17.7	64.3–73.9	1334.5	-4.506	0.001
	2	81.9 \pm 17	78.3–85.5			
BMI	1	25.1 \pm 5.5	23.6–26.6	1233.5	-4.924	0.001
	2	30.4 \pm 5.9	29.1–31.7			
WC	1	82 \pm 11.5	78.9–85.1	1333.5	-4.511	0.001
	2	90 \pm 8.7	88.2–91.9			
TC	1	5.5 \pm 1.1	5.2–5.8	2204.5	-0.894	0.371
	2	5.7 \pm 0.9	5.5–5.9			
LDL-C	1	3.2 \pm 0.9	2.9–3.4	2133	-1.191	0.234
	2	3.4 \pm 0.9	3.2–3.6			
HDL-C	1	1.4 \pm 0.3	1.3–1.5	1791	-2.610	0.009
	2	1.2 \pm 0.3	1.1–1.3			
TG	1	1.5 \pm 0.5	1.4–1.7	1823	-2.477	0.013
	2	2 \pm 0.9	1.8–2.2			
Glucose	1	5.5 \pm 0.7	5.3–5.7	2094	-1.353	0.176
	2	5.8 \pm 1.2	5.5–6.1			

Note: 1 – H. pylori-negative (n=55); 2 – H. pylori-positive (n=88).

Data analysis concludes that GSD patients with MetS are highly infected with H. pylori (61.5 % versus 38.5 %). In their study, S. B. Pradhan et al. wrote about a total of

necrosis factor- α , interleukins (IL-1, IL-6, IL-8, IL-10) eicosanoids. They can act remotely from *H. pylori*'s natural habitat [15] and participate in the pathogenesis of MetS, its components, and cholelithiasis.

The statistically significant relationship between *H. pylori* and obesity that we found correlates with other researchers results [6]. A study by Li-Wei Chen et al., which included 2604 subjects, showed *H. pylori*-positive effect on obesity development [16]. So, *H. pylori* affects producing ghrelin and leptin, which are involved in metabolic and energy balance regulation and are responsible for hunger and satiety formation. Studies conducted have shown a decrease in leptin and ghrelin serum levels in patients infected with *H. pylori*. Leptin forms a feeling of fullness and suppresses food intake, and its decrease is associated with excessive food consumption and obesity degree. Moreover, a decrease in ghrelin concentration in blood plasma is a physiological adaptation to a positive energy balance associated with excess body weight. CagA protein is also involved in the formation of obesity in individuals with *H. pylori* infection. Thus, according to one of the studies, in CagA-positive patients infected with *H. pylori*, BMI was higher than in CagA-negative ones [1.13+/-0.26 mm vs 0.97+/-0.15 mm; univariate analysis, $p=0.0001$; multivariate analysis: OR=2.36; 95 % CI=1.57–3.54; $p=0.0001$] [17]. Also, in overweight individuals, changes in innate and adaptive immunity such as slowing monocyte and macrophage maturation and decreased polymorphic-nuclear bactericidal ability are observed. This undoubtedly increases *H. pylori* colonising the gastric mucosa. In obese patients, natural killer cells (NK) metabolic reprogramming that violates immunological surveillance is detected. Intestinal microbiota changes caused by *H. pylori* also contribute to patient overweight formation.

Our data indicate an increase in the frequency of DM and elevated glucose levels in patients with *H. pylori* infection. Similar results were recorded in other studies [6]. Interestingly, patients with *H. pylori* infection, especially young people, have defects in insulin secretion associated with an increase in gastrin concentration and a decrease in serum somatostatin levels. This leads to a decrease in insulin levels, followed by carbohydrate tolerance formation and DM development. It is worth noting that *H. pylori* infection is closely linked with a decrease in the pepsinogen I/pepsinogen II ratio associated with serum glucose levels. DM also causes cellular and humoral immunity violation, which increases vulnerability to *H. pylori* infection. An imbalance in carbohydrate metabolism inhibits gastrointestinal tract motility, which results in a delay of infected evacuation from the stomach and a decrease in hydrochloric acid secretion. This, in turn, increases its colonization by pathogenic bacteria. Shifts in the intestinal microbiota associated with *H. pylori* infection correlate with changes in carbohydrate metabolism [14]. With microbial dysbiosis, there is an increase in the harmful metabolite number and a change in the bile acid composition, which comes from carbohydrate and protein fermentation. As a result, insulin resistance pathways are activated and initiate obesity, diabetes, and atherosclerosis [14].

Epidemiological studies have shown that *H. pylori* infection is associated with cardiovascular diseases and their risk factors. Z. Wan et al. conducted a cross-sectional study, which included 5,246 adult participants, and found that *H. pylori* infection correlated with AH prevalence [18]. Also, there is evidence that patients with *H. pylori* infection are prone to atherosclerotic vascular lesions caused by changes in the lipid profile.

Among our study subjects, patients infected with *H. pylori* had severe dyslipidemia compared with uninfected ones. We have established a statistically significant relationship between *H. pylori* infection, decreased HDL, and increased serum TG. Similar results were obtained by other authors [6]. A. Hoffmeister et al. also recorded lower HDL-C and WC values in infected patients ($p=0.005$) [19]. T. Gunji et al., in addition to the positive association between *H. pylori* and low HDL-C values ($p<0.001$), found a link with LDL-C ($p=0.005$) [20] A. Laurila et al. [21] reported a significant TG increase in *H. pylori*-positive patients [10]. These changes in lipid homeostasis were significant even after adjustment of the socioeconomic status, body weight, age, and DM [21].

H. pylori infection affects cytokine production, including IL-6 release [15]. Exciting data are presented in a study by Pohjanen et al., which compared serum lipids concentration in *H. pylori*-positive and negative patients and controlled IL-6-174 polymorphism. As a result, data were obtained proving that *H. pylori* infection is more associated with low HDL levels in the serum of patients with the IL-6 -174 genotype. This suggests that the association between *H. pylori* infection and serum HDL can be transmitted through IL-6 [22]. Thus, these immune system reactions provoke changes in lipid metabolism and even contribute to coronary heart disease development. In addition, between the reduced pepsinogen I / pepsinogen II ratio resulting from *H. pylori* infection there is a linear relationship with TG ($P<0.05$), HDL-C ($P<0.01$), the TG/HDL ratio ($p<0.05$), and an increase in the lipid accumulation product index [23].

Successful *H. pylori* eradication therapy has a beneficial effect on metabolic processes, improving the lipid profile, decreasing insulin resistance, blood pressure, and contributing to a decrease in the pro-inflammatory cytokine level. *H. pylori* has mechanisms of protection against host immunity and antibiotic therapy, which forces researchers worldwide to seek new treatments for the infection. Due to a strong connection between cholelithiasis, *H. pylori*, and metabolic disorders. Some scientists are considering GSD treatment with ursodeoxycholic acid (UDCA) combined with statins: 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors. This combination normalizes lipid profile indices, increases bile lithogenicity, prevents cholesterol gallstone formation and promotes stone lysis [24]. At the same time, statins have antibacterial activity and act synergistically with antibiotics. Statins reduce *H. pylori* infection risk by reducing the macrophage infection burden. Due to cellular cholesterol depletion under the influence of statins, CagA-induced damage is attenuated [25] and autophagy is enhanced, which undoubtedly has a beneficial effect on immune defense against pathogens.

Thus, in our study, the antibacterial eradication therapy effect was higher in the group of patients taking rosuvastatin compared with those not taking it (95 % versus 80 %), which proves the additive, adjuvant effect of rosuvastatin on *H. pylori* eradication [24]. Therefore, the above data show the relevance of UDCA combination with statins together with antibacterial eradication therapy in patients with GSD and *H. pylori*.

Conclusions. Thus, the results obtained suggest that *H. pylori* promotes gallbladder calculi development by affecting the patients' metabolic profile, acting as a GSD trigger factor. In this regard, a complex of preventive measures, including *H. pylori* eradication, can help reduce the incidence of GSD and GSD-associated diseases of internal organs.

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References

- Rodriguez S., Gaunt T. R., Guo Y., Zheng J., Barnes M. R. [et al.] Lipids, obesity and gallbladder disease in women: Insights from genetic studies using the cardiovascular gene-centric 50K SNP array. *European journal of human genetics: EJHG*. 2015;24:106-112. <https://doi.org/10.1038/ejhg.2015.63>
- Cen L., Pan J., Zhou B., Yu C., Li Y. [et al.] Helicobacter Pylori infection of the gallbladder and the risk of chronic cholecystitis and cholelithiasis: A systematic review and meta-analysis. *Helicobacter*. 2018;23(1). <https://doi.org/10.1111/hel.12457>
- Zhou D., Zhang Y., Gong W., Mohamed S. O., Ogbomo H. [et al.] Are Helicobacter pylori and other Helicobacter species infection associated with human biliary lithiasis? A meta-analysis. *PLoS One*. 2011;6(11):e27390. <https://doi.org/10.1371/journal.pone.0027390>
- Chang W.-L., Yeh Y.-C., Sheu B.-S. The impacts of H. pylori virulence factors on the development of gastroduodenal diseases. *J. Biomed. Sci.* 2018;25. <https://doi.org/10.1186/s12929-018-0466-9>
- Jeyamani L., Jayarajan J., Leelakrishnan V., Swaminathan M. CagA and VacA genes of Helicobacter pylori and their clinical relevance. *Indian J. Pathol. Microbiol.* 2018;61(1):66-69. https://doi.org/10.4103/IJPM.IJPM_234_17
- Upala S., Jaruvongvanich V., Riangwiwat T., Jaruvongvanich S., Sanguankeo A. Association between Helicobacter pylori infection and metabolic syndrome: a systematic review and meta-analysis. *J. Dig. Dis.* 2016;17(7):433-440. <https://doi.org/10.1111/1751-2980.12367>
- Pradhan S. B. Study of Helicobacter hepaticus in gallbladders with cholelithiasis and its sensitivity pattern. *Kathmandu Univ. Med. J. (KUMJ)*. 2009;7:125-128. <https://doi.org/10.3126/kumj.v7i2.2704>
- Yonezawa H., Osaki T., Hojo F., Kamiya S. Effect of Helicobacter pylori biofilm formation on susceptibility to amoxicillin, metronidazole and clarithromycin. *Microb. Pathog.* 2019;132:100-108. <https://doi.org/10.1016/j.micpath.2019.04.030>
- Madsen J. S., Burmølle M., Hansen L. H., Sørensen S. J. The interconnection between biofilm formation and horizontal gene transfer. *FEMS Immunol. Med. Microbiol.* 2012;65(2):183-195. <https://doi.org/10.1111/j.1574-695X.2012.00960.x>
- Wong E. H., Ng C. G., Chua E. G., Tay A. C., Peters F. [et al.] Comparative Genomics Revealed Multiple Helicobacter pylori Genes Associated with Biofilm Formation In Vitro. *PLoS One*. 2016;11(11):e0166835. <https://doi.org/10.1371/journal.pone.0166835>
- Wei J., Noto J. M., Zaika E., Romero-Gallo J., Piazuelo M. B. [et al.] Bacterial CagA protein induces degradation of p53 protein in a p14ARF-dependent manner. *Gut*. 2015;64(7):1040-1048. <https://doi.org/10.1136/gutjnl-2014-307295>
- Li N., Tang B., Jia Y. P., Zhu P., Zhuang Y. [et al.] Helicobacter pylori CagA Protein Negatively Regulates Autophagy and Promotes Inflammatory Response via c-Met-PI3K/Akt-mTOR Signaling Pathway. *Front. Cell. Infect. Microbiol.* 2017;7:417. <https://doi.org/10.3389/fcimb.2017.00417>
- Wunder C., Churin Y., Winau F., Warnecke D., Vieth M. [et al.] Cholesterol glucosylation promotes immune evasion by Helicobacter pylori. *Nat. Med.* 2006;12(9):1030-1038. <https://doi.org/10.1038/nm1480>
- Altuntaş Y., Batman A. Microbiota and metabolic syndrome. *Türk Kardiyol. Dern. Ars.* 2017;45(3):286-296. <https://doi.org/10.5543/tkda.2016.72461>
- Tsai H.-F., Hsu P.-N. Interplay between Helicobacter pylori and immune cells in immune pathogenesis of gastric inflammation and mucosal pathology. *Cell. Mol. Immunol.* 2010;7(4):255-259. <https://doi.org/10.1038/cmi.2010.2>
- Chen L.-W., Kuo S.-F., Chen C.-H., Chien C.-H., Lin C.-L. [et al.] A community-based study on the association between Helicobacter pylori Infection and obesity. *Sci. Rep.* 2018;8(1):10746. <https://doi.org/10.1038/s41598-018-28792-1>
- Diomedes M., Pietroiusti A., Silvestrini M., Rizzato B., Cupini L. M. [et al.] CagA-positive Helicobacter pylori strains may influence the natural history of atherosclerotic stroke. *Neurology*. 2004;63(5):800-804. <https://doi.org/10.1212/01.wnl.0000138025.82419.80>
- Wan Z., Hu L., Hu M., Lei X., Huang Y. [et al.] Helicobacter pylori infection and prevalence of high blood pressure among Chinese adults. *J. Hum. Hypertens.* 2018;32(2):158-164. <https://doi.org/10.1038/s41371-017-0028-8>
- Hoffmeister A., Rothenbacher D., Bode G., Persson K., März W. [et al.] Current Infection with Helicobacter pylori, but not seropositivity to Chlamydia pneumoniae or cytomegalovirus, is associated with an atherogenic, modified lipid profile. *Arterioscler. Thromb. Vasc. Biol.* 2001;21(3):427-432. <https://doi.org/10.1161/01.atv.21.3.427>
- Gunji T., Matsuhashi N., Sato H., Fujibayashi K., Okumura M. [et al.] Helicobacter pylori infection is significantly associated with metabolic syndrome in the Japanese population. *Am. J. Gastroenterol.* 2008;103(12):3005-3010. <https://doi.org/10.1111/j.1572-0241.2008.02151.x>
- Laurila A., Bloigu A., Näyhä S., Hassi J., Leinonen M. [et al.] Association of Helicobacter pylori infection with elevated serum lipids. *Atherosclerosis*. 1999;142(1):207-210. [https://doi.org/10.1016/s0021-9150\(98\)00194-4](https://doi.org/10.1016/s0021-9150(98)00194-4)
- Pohjanen V.-M., Koivurova O.-P., Niemelä S. E., Karttunen R. A. [et al.] Role of Helicobacter pylori and interleukin 6-174 gene polymorphism in dyslipidemia: a case-control study. *BMJ Open*. 2016;6(1):e009987. <https://doi.org/10.1136/bmjopen-2015-009987>
- Bahadoran Z., Mirmiran P., Zarif-Yeaganeh M., Zojaji H., Azizi F. Helicobacter pylori Stool Antigen Levels and Serological Biomarkers of Gastric Inflammation are Associated with Cardio-Metabolic Risk Factors in Type 2 Diabetic Patients. *Endocrinol. Metab. (Seoul)*. 2015;30(3):280-287. <https://doi.org/10.3803/EnM.2015.30.3.280>
- Osadchuk M. A., Mironova E. D., Vasileva, I. N., Korzhnikov N. P., Achkasov E. E. [et al.] Oral medical dissolution of gallstones in patients with gallstone disease. *Pres. Méd. Argent.* 2019;1:34-40.
- Lai C. H., Chang Y. C., Du S. Y., Wang H. J., Kuo C. H. [et al.] Cholesterol depletion reduces Helicobacter pylori CagA translocation and CagA-induced responses in AGS cells. *Infect. Immun.* 2008;76(7):3293-3303. <https://doi.org/10.1128/IAI.00365-08>

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